

## Mucosal healing and inflammatory bowel disease: Therapeutic implications and new targets

Megan Lynn Otte, Raju Lama Tamang, Julia Papapanagiotou, Rizwan Ahmad, Punita Dhawan, Amar B Singh

**Specialty type:** Gastroenterology and hepatology

**Provenance and peer review:** Invited article; Externally peer reviewed.

**Peer-review model:** Single blind

**Peer-review report's scientific quality classification**

Grade A (Excellent): 0  
Grade B (Very good): B  
Grade C (Good): C  
Grade D (Fair): 0  
Grade E (Poor): 0

**P-Reviewer:** Long P, China; Mohammadi S, Iran

**Received:** October 21, 2022

**Peer-review started:** October 21, 2022

**First decision:** November 30, 2022

**Revised:** December 16, 2022

**Accepted:** February 14, 2023

**Article in press:** February 14, 2023

**Published online:** February 21, 2023



**Megan Lynn Otte**, Department of Pathology and Microbiology, University of Nebraska Medical Center, Omaha, NE 68198, United States

**Raju Lama Tamang, Julia Papapanagiotou, Rizwan Ahmad, Punita Dhawan, Amar B Singh**, Department of Biochemistry and Molecular Biology, University of Nebraska Medical Center, Omaha, NE 68198, United States

**Corresponding author:** Amar B Singh, PhD, Professor, Department of Biochemistry and Molecular Biology, University of Nebraska Medical Center, 600 42nd Dewey, Omaha, NE 68118, United States. [amar.singh@unmc.edu](mailto:amar.singh@unmc.edu)

### Abstract

Mucosal healing (MH) is vital in maintaining homeostasis within the gut and protecting against injury and infections. Multiple factors and signaling pathways contribute in a dynamic and coordinated manner to maintain intestinal homeostasis and mucosal regeneration/repair. However, when intestinal homeostasis becomes chronically disturbed and an inflammatory immune response is constitutively active due to impairment of the intestinal epithelial barrier autoimmune disease results, particularly inflammatory bowel disease (IBD). Many proteins and signaling pathways become dysregulated or impaired during these pathological conditions, with the mechanisms of regulation just beginning to be understood. Consequently, there remains a relative lack of broadly effective therapeutics that can restore MH due to the complexity of both the disease and healing processes, so tissue damage in the gastrointestinal tract of patients, even those in clinical remission, persists. With increased understanding of the molecular mechanisms of IBD and MH, tissue damage from autoimmune disease may in the future be ameliorated by developing therapeutics that enhance the body's own healing response. In this review, we introduce the concept of mucosal healing and its relevance in IBD as well as discuss the mechanisms of IBD and potential strategies for altering these processes and inducing MH.

**Key Words:** Inflammation; Injury/repair; Mucosal healing; Mucosal barrier; Therapeutics; Colitis

©The Author(s) 2023. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core Tip:** Mucosal healing (MH) is vital in maintaining intestinal homeostasis and protecting against infection and injury. MH has emerged as an important clinical criterion in effective treatment of inflammatory bowel disease (IBD). However, there remains a relative lack of therapeutics that can restore MH due to the complexity of the disease and healing processes. Through increased understanding of the molecular mechanisms of MH, tissue damage from IBD may be ameliorated by developing novel therapeutics. Here, we introduce the concept of MH and its relevance in IBD, and discuss the mechanisms of IBD and potential strategies for altering these processes for inducing MH.

**Citation:** Otte ML, Lama Tamang R, Papapanagiotou J, Ahmad R, Dhawan P, Singh AB. Mucosal healing and inflammatory bowel disease: Therapeutic implications and new targets. *World J Gastroenterol* 2023; 29(7): 1157-1172

**URL:** <https://www.wjgnet.com/1007-9327/full/v29/i7/1157.htm>

**DOI:** <https://dx.doi.org/10.3748/wjg.v29.i7.1157>

## INTRODUCTION

Mucosal healing (MH) is a process of wound repair that restores the integrity of damaged epithelial barrier and homeostatic function after an injury compromises barrier integrity[1]. It is a complex process regulated by multiple cell types, through distinct mechanisms in response to highly specific stimuli within multiple signaling and cytokine pathways[1]. For simplification, the MH process is considered to have three phases: Epithelial restitution, proliferation, and differentiation and maturation[1]. Restitution consists of epithelial cells migrating into a wound within hours, followed by proliferation of epithelial cells in hours to days, and finally differentiation of intestinal stem cells into all mature intestinal cell types[1-3]. Each phase is induced and regulated by multiple cytokines, growth factors, and cell types reviewed in[1] and can be influenced by many factors that enhance or prevent wound healing as well as by the source of barrier injury. When the homeostatic process of wound healing is slowed or delayed by external or genetic factors, chronic inflammation may develop because repair of the intestinal epithelial barrier (IEB) and subsequent reduction of inflammation will not occur unless wound healing mechanisms are present[3]. Due to inflammation and chronic immune response, the consequences of impaired MH are chronicity of autoimmune diseases, including Inflammatory Bowel Disease, (IBD) and its progression to colorectal cancer.

## INFLAMMATORY BOWEL DISEASE OVERVIEW AND PATHOGENESIS

Inflammatory Bowel Disease (IBD) is a term that represents autoimmune inflammatory diseases of the gut, with ulcerative colitis (UC) and Crohn's Disease (CD) being the major disease types; however, the etiology of IBD remains unclear. IBD is known to have genetic and environmental risk factors, but the mechanisms by which these factors induce IBD are not well-understood[4]. Common symptoms include abdominal pain, diarrhea, weight loss, malnutrition, and particularly in CD, nausea, vomiting, intestinal blockages, fistulae, and abscesses[4,5]. IBD is an increasingly common and often debilitating disease, affecting up to 200 individuals *per* 100000 people in the United States[4]. Onset of IBD often occurs before the age of 30, and patients experience poor quality of life along with high risk of developing colorectal cancer due to the chronic and progressive symptoms and persistent inflammatory state[1,4]. Though multiple treatment mechanisms exist, including corticosteroids, anti-inflammatory medications, monoclonal antibodies, stem cell treatments, and surgery, IBD cannot currently be cured[4]. Therefore, continued research into the mechanisms of pathogenesis and development of new pharmaceuticals and treatment methods is vital to decrease mortality and improve quality of life for IBD patients.

Autoimmune disease pathogenesis is difficult to study due to the multifactorial causes of disease and complex molecular mechanisms, as well as the predicament of patients not presenting to the clinic until late in the disease development process. Although the disease complexity and difficulty in identifying the initial molecular instigators has thus far precluded full understanding of the specific molecular mechanisms of IBD pathogenesis, the general overarching processes have been elucidated. During IBD pathogenesis, the IEB and mucosal layers become damaged and inflamed due to injury and/or infection, which can develop into a state of chronic inflammation and reduced IEB integrity. Genetic factors can also play a role in pathogenesis, particularly in CD through genes such as a variant that impairs autophagy and dysregulates the IEB and gut microbiota[6,7]. The damage to the IEB results in microbial and antigen exposure in the intestinal lumen, leading to an inflammatory cascade and disturbed homeostasis[8,9]. Multiple cytokines and immune cell types are also thought to contribute to the pathogenesis of or protection against IBD; therefore, further study of the complex interactions governing the maintenance or breakdown of gut barrier homeostasis is imperative to deeper

understanding of IBD pathogenesis and the development of effective treatments.

## MUCOSAL HEALING AND PATHOLOGICAL RELEVANCE IN IBD

The main goals of IBD treatment are two-pronged: Reducing symptoms and preventing new inflammation and intestinal injury through traditional treatment methods, and recently a new, ambitious goal of inducing wound healing of existing inflammation and damage[8]. Some current treatments may contribute to both treatment goals; however, many existing clinical treatments are more targeted toward the traditional goal of preventing inflammation and damage. Even with the advent of some newer MH-focused treatments, additional avenues of further enhancing MH should be explored. Although clinical remission or preventing new inflammation is possible, patients may still have residual disease symptoms during remission due to the defective wound healing process leaving previous intestinal damage unrepaired. Additionally, up to half of IBD patients experience non-response or loss of response to standard therapeutics, leading to relapse[10]. Hence, MH induction presents an attractive goal in effective long-term treatment of IBD and prevention of relapse, and thus also prevention of progression to colitis-associated colon cancer. In this review, we will summarize the main current and prospective treatments of IBD, as described in Table 1, and their benefits and limitations towards the goal of reducing inflammation and achieving MH. More detailed analysis of the mechanisms of action, safety, and efficacy profiles of current IBD therapies and clinical trials can be found in Neurath *et al*[11]. However, in this review, we will emphasize areas that have not yet been as extensively clinically explored, including the temporal control of gut immune function, which presents novel potential for fine-tuning of the immune system in MH and restoration of gut homeostasis. We will examine three major factors contributing to the pathogenesis and tissue damage of IBD, as shown in Figure 1: Gut barrier dysfunction, gut dysbiosis, and inflammatory cytokine responses. Specifically, we will focus on the prospect of altering these factors and associated pathways summarized in Table 2 for both the reduction of inflammation and induction of MH.

## CURRENT STATUS OF IBD THERAPIES AND FOCUS ON ENHANCED MUCOSAL HEALING

### **Traditional Therapies**

Historically, there has been a disconnect between the expectation of IBD treatment promoting MH and real treatment outcomes, as many therapies for UC and CD primarily target symptom relief and reduction of chronic inflammation[12]. Corticosteroids, Methotrexate, and surgery are typically utilized to achieve these goals, but they do not promote MH as the primary therapeutic endpoint[13,14]. Because UC and CD are progressive diseases, patients may still experience intestinal damage even during periods without physical symptoms, and disease progression is typically only slowed by these treatments, not stopped[12,15]. However, achieving MH may help stop disease progression as well as decrease symptom severity. New treatment plans broaden the therapeutic focus to include inducing MH through a variety of mechanisms, such as by altering the gut microbiome and altering inflammatory and anti-inflammatory cytokines with antibodies or exogenous cytokine therapies.

### **Inflammation Reduction and Immune Modulation Therapies**

One such mechanism for emphasizing MH includes suppressing specific parts of the patient's immune system to decrease the main contributors of chronic inflammation. When new inflammation is reduced, it may then be possible for the wound healing process to begin to keep up with the rate of tissue damage. Methods of achieving this goal include enteral nutrition (EN), partial EN (PEN), 5-aminosalicylates (5-ASA), and Azathioprine treatments, which focus on decreasing IBD-associated inflammation by suppressing the host immune system in a less global manner than corticosteroids or preventing the production of inflammatory molecules. EN is a dietary treatment for IBD patients that can either totally (with EN) or partially (with PEN) replace solid food intake with specialized formula[16]. Although the mechanism by which EN induces MH is currently unknown, microbiome changes are implicated and are hypothesized to aid in decreasing chronic inflammation[17]. While typically a pediatric treatment, a pilot study shows MH in adults following EN and PEN treatment as well[16]. Currently, the largest drawback of EN treatment is low patient compliance, especially in adults[16]. 5-ASA drugs, which are used to induce remission in early IBD, are shown to induce MH in 43.7% of patients[18]. Azathioprine is a purine analog inhibiting purine metabolism and blocking T cell activation and co-stimulation, therefore functioning by suppressing the immune system and decreasing inflammation in IBD[11,19-21]. It is shown to achieve MH independently in some cases (30.1%) alone but is more successful when used in conjunction with anti-TNF- $\alpha$  antibodies such as Infliximab (44%)[22]. Other immunomodulating drugs shown to induce MH when given with monoclonal antibodies include Cyclosporine and Tacrolimus[23].

**Table 1 Summary of inflammatory bowel disease therapeutics**

Treatment type	Available therapeutics	Mucosal healing relevance/ Success
Corticosteroids	Prednisone/ Prednisolone/ Methylprednisone	Prednisone treatment for 14 d (20 mg/day) decreased mucosal inflammation indicating a possible role in developing short-term MH[139]. 29% of patients in one study displayed endoscopic remission after steroid treatment[140].
Nutritional therapy	Enteral nutrition (EN)/ Partial enteral nutrition (PEN)	EN/PEN induce MH in both adults and children[14].
Aminosalicylates (5-ASA)	Sulfasalazine/ Mesalamine/ Olsalazine/ Balsalazide	On average induce MH in 43.7% of patients[141].
Immunomodulators	Azathioprine/ 6-mercaptopurine	Azathioprine alone has achieved MH in 16.5% of cases and in 43.9% when used in combination with antibody therapies[18]. After 16 wk of mercaptopurine treatment, patients in remission showed a 47.1% rate of MH[142].
	Cyclosporine	Shown to induce MH when used in conjunction with Vedolizumab[143].
	Tacrolimus	Shown to induce MH when used in conjunction with Vedolizumab[143].
	Methotrexate	After 36 wk, methotrexate treatment had a MH rate of 47.4%[142].
Monoclonal antibody/ Biologic therapies	Adalimumab	Induced MH in 24% of patients treated[24].
	Certolizumab	Clinical response rate at weeks 2 and 12 was 29.7% and 52.8% (respectively) in CD[25].
	Infliximab	Treatment induced MH in up to 60.3% of patients in phase 2 clinical trials[23].
	Natalizumab	MH achieved by 42.3% of patients after 14.1 mo of treatment[144].
	Risankizumab-rzaa	Endoscopic response and deep remission observed in 55% and 29% of patients (respectively), indicating MH[27].
	Ustekinumab	Treatment of individuals with moderate to severe CD showed MH <i>via</i> a reduced disease score after 8 wk[19].
	Vedolizumab	Has shown to induce MH in up to 50% of UC patients and 29% of CD patients in clinical trials[26,27].

MH: Mucosal healing; EN: Enteral nutrition; PEN: Partial enteral nutrition; 5-ASA: Aminosalicylates; CD: Chron's Disease; UC: Ulcerative colitis.

### Monoclonal Antibody Therapies

Monoclonal antibodies targeting inflammatory cytokines are an emerging class of IBD therapies that more directly focus on permitting or inducing MH and are an attractive method of IBD treatment. Monoclonal antibody drugs targeting multiple cytokines are approved or in trials, and function by removing inflammatory cytokines. Anti-tumor necrosis factor-alpha (TNF- $\alpha$ ) antibody-based drugs such as Infliximab, Adalimumab, and Certolizumab are biologic therapies that have achieved significant clinical success and are now widely used as front-line treatments for IBD, with the goal of reducing inflammation and promoting MH. The target of anti-TNF- $\alpha$  therapies is a pro-inflammatory cytokine contributing to the chronic and severe inflammatory response observed in UC and CD, and the monoclonal antibody functions by downregulating pro-inflammatory molecules and restoring IEB integrity, allowing the body to begin to heal[8,24-26]. Infliximab is an Immunoglobulin G1 anti-TNF- $\alpha$  antibody that is shown to induce MH. Active phase 1 and 2 clinical trials of Infliximab in UC patients find that, respectively, 45.5% of patients and 60.3% of patients exhibit MH[27]. Adalimumab is another anti-TNF- $\alpha$  antibody presenting some indication of MH effects. In the double blind, randomized, placebo controlled clinical trial, 24% of patients display MH at week 52 compared to 0% in the placebo group[28]. Certolizumab pegol is an anti-TNF- $\alpha$  antibody fragment targeting soluble and transmembrane TNF. Its role in promoting MH is not yet well-investigated, but it demonstrates symptom relief and remission at week 8[29]. In some patients, anti-TNF- $\alpha$  treatments are ineffective or lose efficacy over time. In these patients, biologics targeting other inflammatory cytokines are employed as IBD treatment options. For example, Ustekinumab is a monoclonal antibody that targets interleukin (IL)-12 and IL-23 pro-inflammatory cytokines. Ustekinumab significantly reduces SES-CD (Simplified Endoscopic Activity Score for Crohn's Disease) scores in patients over 44 wk, implying its ability to enhance MH[30]. Similarly, Risankizumab, an IL-23 antibody, induced remission in 45% of patients, with an endoscopic response rate of 29%[31]. Additionally, Natalizumab, an anti- $\alpha_4\beta_7$  integrin antibody, induces MH in 42.3% of patients[32]. Vedolizumab, on the other hand, is an  $\alpha_4\beta_7$  integrin receptor antagonist and biologic medication, with potential immunosuppressive effects localized to the colon. In a clinical trial, 50% of UC patients and 29% of CD patients display MH following long-term use[33,34].

Although inflammation reduction and monoclonal antibody treatments have greatly enhanced the quality of IBD treatment in recent years, lack or loss of response occurs with concerning frequency, and even in patients who respond well to treatments, complete resolution of the disease has not been

Table 2 Summary of molecular pathways involved in mucosal healing

Pathways/ Mechanism of action	Associated models studied	Ref.
EGFR signaling	<i>In vitro</i> , colorectal cancer mice, EGFR mutant mice	[43,116]
Hippo/YAP signaling	<i>In vitro</i> , YAP-1 transgenic mice	[36,59]
Notch signaling	Villin-Claudin-1 transgenic mice	[41,42]
Wnt/ $\beta$ -catenin signaling	<i>In vitro</i> and <i>In vivo</i> models of injury/repair	[44,60,61]
Vitamin D receptor (VDR) signaling	<i>In vitro</i> , VDR knockout mice	[45]
Src/focal adhesion kinase	<i>In vitro</i> , Mechanical colonic wound in mice, Nox1 and AnxA1 knockout mice, oral gavage in mice	[76-78]
Autophagy/ATG16L1	Patient biopsies; ATG16L1 T300A knock-in mice; Atg5-manipulated mice	[6,7,104]
SCFA-mediated signaling [acetate, propionate, butyrate, <i>etc.</i> ]	<i>In vitro</i> , Patient biopsies, oral gavage in mice. T-cell induced colitis, trinitrobenzenesulphonic acid (TNBS) colitis	[83-85,91,93,100,101,114]
TLR-mediated signaling	DSS colitis	[109,110,112]
MyD88 mediated bacterial sensing	Mechanical colonic wound, MyD88 knockout mice	[111]
Prostaglandin-endoperoxidase synthase 2 enzyme (PGE2)	<i>In vitro</i> , mechanical colonic wound, Ptg2 knockout mice, Ptg4 knockout mice	[111,112]
Mucin 2 signaling	<i>In vitro</i> , DSS colitis, EGFR mutant mice	[80,116]
IL-6/IL-22/IL-23/STAT3 signaling	DSS colitis, Th2-mediated colitis, cytokine deficient mice, bone marrow transplant mice, T-cell induced colitis, human and mouse intestinal organoid culture	[94,97,98,136-138]
TGF- $\beta$ signaling	<i>In vitro</i> , DSS colitis, TGF- $\beta$ transgenic mice	[50,130,131]
IL-10 signaling	<i>In vitro</i> , mechanical colonic wound in mice, IL-10-deficient mice	[132,133]

EGFR: Epidermal growth factor receptor; YAP: Yes-associated protein 1; ATG16L1: Autophagy related 16 like 1 protein; Atg5: Autophagy related 5; SCFA: Short chain fatty acid; TNBS: Trinitrobenzenesulphonic acid; TLR: Toll-like receptor; DSS: Dextran sodium sulfate; PGE2: Prostaglandin-endoperoxidase synthase 2 enzyme; IL: Interleukin; STAT3: Signal transducer and activator of transcription 3; TGF- $\beta$ : Transforming growth factor- $\beta$ .

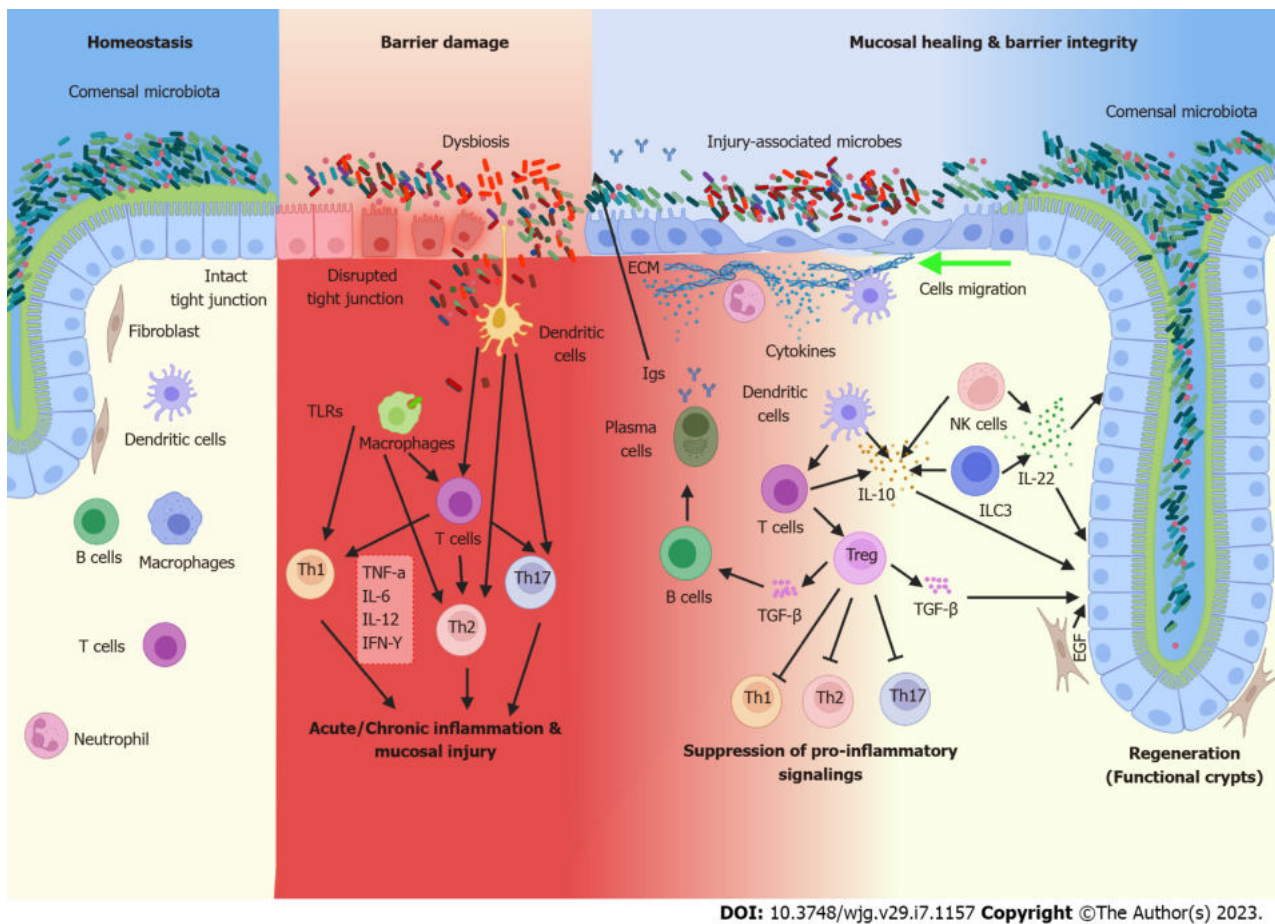
achieved<sup>101</sup>. This is due to the complex nature of disease development through many aberrant proteins and signaling pathways, as well as the multi-faceted process of MH that must be approached from many angles to restore complete IEB homeostasis. Overall, while current therapies offer evidence of permitting MH in IBD, some of these therapies do not directly promote MH through enhancing the processes of restitution, proliferation, or differentiation, but rather by simply inducing immune suppression to decrease inflammation and associated injury. Subsequently, there is a critical need to better understand the processes of inflammation and MH in IBD to aid in the development of actively MH-inducing IBD therapies.

## IBD TISSUE DAMAGE MECHANISMS AND IMPLICATIONS FOR MH INDUCTION

### ***Gut Barrier Functions and Mucosal Healing***

A major role of the intestinal epithelium is its function as a barrier against the luminal environment and antigens; a role that is critical in maintaining normal mammalian homeostasis<sup>[35]</sup>. Accordingly, IEB dysregulation is a major factor in gut inflammation; thus, reinforcement of IEB integrity is a key consideration when developing more effective treatments for gut inflammatory diseases, including IBD. The MH process has been shown to help reinforce barrier integrity<sup>[36]</sup>. A complex and dynamic coordination between epithelial and immune factors facilitates MH, however, the ‘paradoxical’ role of the barrier-integral proteins in promoting MH remains ill-understood. Here, we summarize the key components of IEB regulation to illustrate the dynamic causal relationship between MH and the proteins comprising the structural and functional units of the gut barrier.

The physical component of the IEB consists of a single layer of epithelium, with cells linked by junctional complexes along the apicobasal axis. Excellent reviews have described the types and roles of junctional complex proteins in barrier integrity, thus here we focus on mechanisms by which regulation of these proteins influences inflammation and MH<sup>[35-39]</sup>. Tight junctions, the most apical cell-cell adhesions are considered the “gate” of the IEB and consist of multiple proteins, including the Claudin family of proteins, Occludin, junctional adhesion molecule (JAM) and the zonula occludens (ZO)-proteins<sup>[40]</sup>. Studies in cell and mouse models demonstrate through genetic manipulation that tight



DOI: 10.3748/wjg.v29.i7.1157 Copyright ©The Author(s) 2023.

**Figure 1** Pictorial depiction of inter-connection between the immune system, inflammation, and microbiota in mucosal inflammation, associated injury, and healing. Left: normal mucosal homeostasis; Middle: Inflammatory lesions damage the mucosal barrier between the gut lumen and the rest of the body. Barrier damage leads to immune cell activation, cytokine release, and feedback cycles of deteriorating inflammation driven by microbes crossing the damaged barrier; Right: Migration of circulating restitutive immune cells to the wound area, the release of repairing cytokines; crosstalk among extracellular matrix and epithelial cells for proliferation and migration; switching of microbiota and cytokines for mucosal healing and functional crypt regeneration.

junction proteins are not merely static structural entities of the IEB; they perform additional non-canonical roles of regulating epithelial cell proliferation, survival, differentiation, and migration, which are the same processes integral to MH. In this regard, ZO-1 regulates expression and nuclear localization of the transcription factor ZO-1-associated nucleic-acid-binding protein to influence cell proliferation in a density dependent manner[41,42]. Recent studies also show that the ZO-proteins modulate Hippo/Yes-associated protein 1 signaling, a critical regulator of crypt growth and MH[43]. A similar role of JAM in regulating intestinal epithelial cell (IEC) proliferation has been reported[44,45]. Occludin, on the other hand, is shown to regulate IEC apoptosis and survival[46,47]. Recent studies demonstrate that the Claudin family of proteins is integral to tight junction structure and function and plays a complex role in gut inflammation and regenerative processes. For example, Claudin-1 overexpression in the intestinal epithelium of mouse models of IBD induces significant dysregulation of Notch/Wnt signaling and severe colitis, resulting in delayed recovery from colitis-associated injury[48, 49]. Claudin-2 is unique among the Claudin proteins expressed in the intestine, as it is primarily expressed at the crypt base among proliferative undifferentiated cells and associates inversely with the differentiation state of IECs[48]. Of note, Claudin-2 is a direct target of Epidermal Growth Factor Receptor (EGFR), Wnt/ $\beta$ -catenin, and Vitamin D receptor signaling, all of which promote intestinal MH [50-52]. Claudin-3 on the other hand, which functions as a receptor for *Clostridium perfringens*, is sharply downregulated in the biopsies of the IBD patients, and loss of its function in the mouse gut promotes colitis-associated cancer[53-55]. We have also previously reported that Claudin-3 Loss enhances gp130/IL-6/STAT3 (signal transducer and activator of transcription 3) signaling, which promotes colitis-associated injury/repair[50]. Interestingly, the loss of intestinal Claudin-7 expression results in spontaneous inflammation due to dysregulation of epithelial-extracellular matrix interactions[56,57]. Claudin-7 also regulates intestinal stem cell function in association with the Epithelial Cellular Adhesion Molecule protein[58,59]. Conversely, Claudin-15 Loss in the IEB results in a mega-intestine[60]. Hence, dysregulation of tight junction protein expression in the IEB results in IBD, indicating that restoration of tight junction homeostasis is a vital component of MH and a promising target for treatment

development.

Like tight junction component proteins, adherens junction proteins contribute significantly to gut inflammation and MH. Here, E-cadherin, a protein whose expression indicates an epithelial phenotype, contributes to inflammation-associated epithelial repair by regulating the epithelial-to-mesenchymal transition, a process associated with cell proliferation and migration[58,61,62]. Specifically, E-cadherin expression inhibits migration of IECs and wound healing[63]. Contrastingly, complete E-cadherin loss causes a severe inflammatory phenotype characterized by villus blunting, which is a marker of premature epithelial death, and incomplete brush border development[64]. The stabilization of E-cadherin expression is facilitated partly by binding with the cytoplasmic domain of p120-cadherin. Accordingly, p120 knockout mice displayed disrupted intestinal integrity and early death from intestinal injury[65]. Other heterodimeric adherens junction proteins,  $\alpha$ - and  $\beta$ -catenin, are key players in the regulation of Hippo and Wnt signaling[66,67]. Although  $\alpha$ - and  $\beta$ -catenin expression dictates IEC proliferation and differentiation during injury repair, their expression counter-acts each other. Moreover, ubiquitination of  $\beta$ -catenin by  $\alpha$ -catenin aids the degradation of  $\beta$ -catenin, balancing the Wnt signaling pathway[68]. Taken together, these findings support a complex and dynamic interdependence between gut barrier regulation and MH, which should be considered for therapeutic potential.

### **Gut Dysbiosis, IBD, and Mucosal Healing**

Aberrant microbiome-immune interactions can lead to improper immune activation and are potentially responsible for the clinical and endoscopic observations in IBD patients. Mechanisms of microbial involvement in IBD include production of short-chain fatty acids (SCFAs), interaction with autophagy pathways, activation of immune cells, TLR signaling, and prostaglandin pathways[69-73]. Excellent reviews have covered the details of such interactions and the dynamic association with gut inflammatory processes[71-75]. Thus, here we focus primarily on how gut microbiota may contribute to MH processes under normal and/or inflammatory conditions.

Gut dysbiosis is mediated by pathogenic microbes harboring genes encoding toxin proteins, which disrupt the IEB *via* disassembly or redistribution of tight junction proteins. For example, human epithelial cells treated with *Escherichia coli* or *Salmonella typhimurium* demonstrate downregulation of ZO-1 and Occludin proteins while by contrast, Claudin-2 is upregulated[76-78]. *Shigella flexneri* and *Campylobacter jejuni* are involved in deregulating E-cadherin, as well as activating IL-8 and nuclear factor kappa-light-chain-enhancer of activated B cells (NF- $\kappa$ B), thereby inducing barrier dysfunction and inflammation[79,80]. Overall, studies suggest that pathogenic gut bacteria promote chronic mucosal inflammation by dysregulating IEB integrity.

On the other hand, commensal gut bacteria seem to promote the initial stage of epithelial restitution, as studies in germ-free mice show impaired rates of epithelial cell migration, which is dependent on the formation of focal adhesions[81,82]. In this context, a commensal bacterium activates the focal adhesion kinase, thereby enhancing epithelial restitution and promoting mucosal wound repair in a redox-dependent manner[83-85]. In a mouse colonoscopy-based wound healing model, an abundance of anaerobic bacteria such as *Akkermansia spp* augmented early stages of MH[86]. The selection of mucin-producing bacteria from the mucin layer also helps close mucosal wounds[87,88]. These microbes help generate SCFAs such as acetate, propionate, and butyrate, which are considered the primary energy sources of gut colonocytes and are therefore critical supporters of IEB restoration and integrity following tissue damage[89-92]. The major producers of SCFAs include the genus *Bacteroides*, *Clostridium* clusters IV and XIVa, and *Bifidobacterium*, though they use diverse mechanisms to achieve homeostatic outcomes[89,93,94]. For example, *Bacteroides ovatus* decreases lipopolysaccharide-induced inflammation and produces indole-3-acetic acid that likely promotes IL-22 production by immune cells, yielding beneficial effects in epithelial regeneration[95,96]. SCFAs produced by fiber-fermenting commensal microbes are also linked to upregulation of Foxp3<sup>+</sup> T regulatory (Tregs) cell development, which have a widely documented role in protection against epithelial injury and colitis [97]. Inhibition of histone deacetylases and/or activation of the latent form of transforming growth factor- $\beta$  (TGF- $\beta$ ) to act as a potent inducer of Tregs are potential mechanisms of SCFAs[98,99]. SCFAs also mediate activation of STAT3 which plays a vital role in mucosal homeostasis[100,101]. *Clostridia*-related segmented filamentous bacteria promote IL-23 production by antigen-presenting cells, which activate type 3 innate lymphoid cells (ILCs) to initiate an IL-23R/IL-22/STAT3 loop, thereby producing serum amyloid A which promotes IL-17 production by Th17 cells[102-105]. The importance of the SCFA propionate is the augmentation of dendritic cell and macrophage hematopoiesis precursors that impact intestinal immunity to control the growth of invading mucosal pathogens[106,107]. A breach in this regulation is a central mechanism in triggering, maintaining, and exacerbating IBD. Supplementation with another important SCFA, butyrate, rescues deficiencies in mitochondrial respiration and increases autophagy in the colonocytes of germ-free mice compared to conventionally raised mice[108]. Due to the critical role in repair of gut dysbiosis, the regulation of specific gut bacteria and SCFAs may therefore possess significant potential for clinical treatment of IBD.

As introduced above, accumulating evidence suggests a causal interdependence between autophagic flux in the intestinal epithelium and gut microbiota colonization. In this regard, CD patients who are homozygous for the ATG16L1<sup>T300A</sup> gene variant exhibit higher abundance of Enterobacteriaceae, Bacteroidaceae and Fusobacteriaceae in the inflamed ileum compared with patients homozygous for the wild

type ATG16L1 allele[6]. Similar findings have been obtained from mice where expression of the autophagy gene Atg7 is genetically knocked out in the gut epithelium, as these mice display altered microbial composition with enrichment in *Clostridium septum*, *Eubacterium cylindroides*, and *Bacteroides fragilis* compared to wild type mice[109]. ATG16L1<sup>T300A</sup> variant mice also show changes in fecal microbiota composition compared to wild type mice, displaying an increase in the order Bacteroidales, which is associated with increased Th17 and Th1 cells in the colon and ileum lamina propria without the development of intestinal inflammation[7]. However, Atg5 deficient mice display reduced bacterial diversity, as observed in IBD patients, and contain a low number of the Lachnospiraceae, Ruminococcaceae, and Akkermansia families that control inflammatory responses[110]. Of note, a role of autophagy in regulating intestinal stem cell function and mucosal injury/repair has been demonstrated by several recent studies[111,112]. Taken together, these studies highlight a complex causal integration between host cell autophagy processes and intestinal microbial communities in regulating intestinal homeostasis and injury/repair.

Additionally, infiltrating immune cells such as macrophages and neutrophils responding to gut dysbiosis comprise essential components of intestinal wound healing by altering aberrant physiological parameters of the local microenvironment, such as microbe-associated molecular patterns (MAMPs) and decreased oxygen levels from the formation of reactive oxygen species[86,113-115]. Of note, Toll-like receptors (TLRs) expressed on multiple immune cell lineages induce signaling pathways upon binding by MAMPs and improve outcomes in experimental mouse colitis models *via* the promotion of wound healing[116,117]. Specific microbes in proximity to the wound bed also activate host epithelial proliferative signaling through a formyl peptide receptor pathway[83,84]. Studies employing a mechanical colonic wound method further disclose a protective role for prostaglandin E2 (PGE2) in re-establishment of the IEB through a TLR2/MyD88-dependent manner[118,119]. A follow-up study from the same group shows that in the early repair phase, a TLR2/PGE2 axis is required for barrier establishment; however, PGE2 must then decrease to allow for epithelial proliferation and regeneration [118]. In this context, Jain *et al*[120] elegantly demonstrates that temporal regulation of the bacterial metabolite PGE2/deoxycholate during colonic repair is critical for crypt regeneration[120]. The highly specific and time-dependent switching of microbial colonization and signaling pathways can therefore act to promote MH in a localized manner.

Several therapeutic approaches have been examined by administration of prebiotics or probiotics to regulate the microbiota. For example, butyrate enemas are effective in treating experimental colitis and UC patients[121,122]. Also, p40, a protein produced by *Lactobacillus rhamnosus* GG (LGG), activates host epithelial EGFR signaling and mediates wound healing[123]. Of note, the mechanism of MH promotion by LGG is *via* a positive effect on epithelial barrier maturation by upregulation of Claudin-3[124]. Recently, genetically modified probiotic bacteria-based precision delivery of human EGF also appears to be a promising intervention against mucosal inflammation through crypt-derived MH and barrier restoration[125,126]. Firmicutes, such as *Faecalibacterium prausnitzii* play an essential role in mucosal barrier homeostasis by regulating NF- $\kappa$ B activation and IL-8 production[125,126]. In another study, oral gavage with *Faecalibacterium prausnitzii* during dextran sodium sulfate (DSS) colitis improves outcomes compared to mice treated with DSS alone, likely due to participation of Claudin-1 and Claudin-2[125]. The probiotic mixture known as VSL #3, containing 4 strains of *Lactobacilli*, 3 strains of *Bifidobacteria*, and one strain of *Streptococcus* is effective in preventing pouchitis and in treating UC flareups[127]. This probiotic functions by partially upregulating mucin production and restoring the IEB by stimulating ZO-1 and Occludin expression while decreasing Claudin-2[127]. Taken together, a complex interdependence exists between gut microbiota and MH processes in the promotion of barrier integrity that should be fully explored for its intriguing potential in improving clinical outcomes.

### **Inflammatory Cytokines in IBD and Mucosal Healing**

Due to dysregulation of many cytokines and growth factors in IBD and the regulatory importance of many of these same molecules in MH, we discuss the potential of altering immune signaling and inflammatory cascades in restoring proper intestinal homeostatic balance. Increasing knowledge of the coordination of these pathways will contribute to the development of more effective and targeted therapies to ameliorate disease while preserving essential immune system function.

Major cytokines and growth factors that are considered pro-inflammatory in IBD include IL-1 $\beta$ , interferon-gamma (IFN- $\gamma$ ), TNF- $\alpha$ , and IL-6. During an infection in the gut, IL-1 $\beta$ , TNF- $\alpha$ , and IFN- $\gamma$  are shown to be produced by inflammatory monocytes, among which IL-1 $\beta$  and TNF- $\alpha$  are associated with increased IEB permeability[2]. TNF- $\alpha$  and IFN- $\gamma$  are also produced by ILCs and serve to recruit and activate additional inflammatory cells[3]. TNF- $\alpha$  is particularly well-studied and often targeted in the treatment of autoimmune diseases as detailed in our discussion of monoclonal antibody therapies above. Although necessary for innate immune responses against acute pathogens and acute DSS colitis, when produced chronically by T cells TNF- $\alpha$  can be a major contributing factor to the loss of epithelial barrier and development of autoimmune disease[128-130]. IFN- $\gamma$  has been shown to be regulated during wound healing of skin epithelium by Tregs, where lack of Tregs resulted in increased IFN- $\gamma$ , accumulation of pro-inflammatory macrophages, and hindered wound healing[131]. Gut microbiota can also impact immune system activation through cytokine signaling. Kuhn *et al.*[132] demonstrates that intraepithelial lymphocytes (IELs) must interact with commensal Bacteroidales order microbes to produce IL-



6 in response to acute *C. rodentium* colitis infection, aiding in repair of the IEB *via* increased Claudin-1 expression[132]. Despite the necessary effects of pro-inflammatory cytokines for immune system response and homeostasis, each also possesses drawbacks. When the location, amount, or duration of cytokine production becomes dysregulated, chronic inflammation and disease can result. IFN- $\gamma$ , TNF- $\alpha$ , and IL-6 are upregulated after secretion by immune cells in the chronic inflammatory state associated with IBD[3,130,132]. These cytokines are known to increase gut permeability by altering tight junction protein expression[132-134]. In particular, increased IL-6 in the IECs and lamina propria mononuclear cells increases Claudin-2, which promotes intestinal permeability and is known to be upregulated in IBD[134,135]. Interestingly, it is IELs that produce IL-6 in a protective manner during acute infection through the c-Jun N-terminal Kinase pathway, rather than the Mitogen-activated protein kinase/extracellular signal-regulated kinase pathway, suggesting both duration and cell type-specific layers of complexity in the role of IL-6[132,134]. It is likely that similar multi-layer and highly context-specific pathways exist for other inflammatory cytokines as well. Therein lies the challenge of harnessing pro-inflammatory cytokines and signaling pathways in the immune system for MH: Targeting modifications toward the decrease of detrimental chronic effects without impairing their beneficial and homeostatic functions. Continuing in the discussion of IL-6, global reduction of IL-6 expression can decrease chronic inflammation in IBD and enhance MH, but may also increase susceptibility to infection[132,136]. It would therefore be ideal to develop a method of reducing IL-6 production in IECs while leaving expression in IELs intact to avoid increasing the risk of dangerous infections in patients. The potential in fine-tuning inflammatory cytokine expression to promote MH is great, but much work remains to ensure it can be accomplished safely without severe detrimental effects to other functions of the immune system.

Conversely, several other cytokines, growth factors, and cell types function primarily in an anti-inflammatory role in IBD and intestinal homeostasis and offer additional mechanisms to enhance MH by resolving chronic inflammation. Growth factors and cytokines considered to be anti-inflammatory include TGF- $\beta$ , IL-10, IL-22, and IL-17. TGF- $\beta$  is a well-studied growth factor that has been demonstrated by Beck *et al*[137], to play a role in the restitution, or IEC migration, phase of MH, evidenced by the lack of IEC migration and impaired wound healing after TGF- $\beta$  inhibition in DSS colitis[8,137]. Additionally, TGF- $\beta$  when produced by a macrophage secretome called SuperMApo aids in removal of apoptotic cells and resolution of inflammation in IBD models[57,138]. Importantly, since this secretome produces TGF- $\beta$  from a singular cell type, rather than globally, its administration can provide the context- and location-dependent production of beneficial TGF- $\beta$  while avoiding potential opposing or off-target effects. Macrophages and Th2 cells also produce IL-10, which enhances the proliferation phase of MH, maintains immune tolerance to the many antigens encountered by the IEB, and promotes barrier integrity[139,140]. Similarly, the production of IL-17 by anti-inflammatory Tregs helps with IEC proliferation and blockage of detrimental microbiota in colitis models, providing protection against IBD[141]. One of the major players in protection against IBD is IL-22, which is produced by multiple cell types and promotes MH by more than one mechanism, as reviewed in[142]. Most importantly in IBD, IL-22 is produced by ILCs, CD4+ T cells, and NK cells, demonstrating activation of both the innate and adaptive immune system[143,144]. IL-22 primarily acts on intestinal stem cells (ISCs) and IECs and functions by activating STAT3 signaling, which induces IEC proliferation and therefore MH[101,142,143,145]. Regarding ISCs, IL-22 both protects them from depletion during intestinal inflammation and induces regeneration[101,143,145]. Though some of these cytokines and growth factors can be inflammatory under certain circumstances, in the context of IBD they function in an anti-inflammatory manner and are beneficial in promoting MH. Some cytokines even demonstrate potential to be produced or administered selectively in only beneficial locations or cell types. Therefore, anti-inflammatory cytokines and growth factors present promising options for treatment of IBD through selectively reducing inflammatory signals and inducing or enhancing the MH process.

---

## CONCLUSION

In recent years, IBD has become increasingly common in the United States and abroad. As progressive and debilitating diseases, UC and CD have long-term negative implications on health. In the past, treatments have focused on reducing the clinical manifestations of the disease, often leaving underlying disease mechanisms active in the gut[12,15]. With increased understanding of the mechanisms and complex pathogenesis of IBD, further innovation in treatment approaches must occur to improve patients' long-term outcomes. Subsequently, instead of merely hoping to reduce symptoms, doctors now desire therapies that actively aid in healing and regeneration of damaged tissue. MH has therefore emerged as the main goal for research and treatment end points to ensure long-term remission, survival, and a good quality of life for patients. Going forward, understanding the interactions regulating the breakdown and regeneration of the IEB, as well as overarching gut homeostasis processes, will be paramount to treating and curing patients with IBD. Monoclonal antibody therapies offer a promising start to revolutionizing treatment, aiming not only to reduce clinical manifestations, but also to interrupt disease activity on a cellular and molecular level. However, even newly developed antibody therapies

cannot by themselves completely resolve IBD and restore total gut homeostasis. Therefore, the three approaches to targeting the molecular machinery governing IBD of restoring the IEB, regulating the gut microbiota, and altering the cytokine signaling-mediated immune response are all being studied as potential mechanisms for achieving MH. Optimal future treatment protocols for IBD will ideally include a combination of these approaches, with the intent of restoring intestinal homeostasis by balancing expression of multiple proteins and repairing several of the many dysregulated pathways involved in IBD pathogenesis.

## FOOTNOTES

**Author contributions:** Otte ML, Lama Tamang R, Papapanagiotou J, and Ahmad R reviewed literature for the latest advances in the field; Otte ML, Lama Tamang R, Papapanagiotou J, Ahmad R, Dhawan P, and Singh AB wrote and revised this paper.

**Conflict-of-interest statement:** All the authors report no relevant conflicts of interest for this article.

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

**Country/Territory of origin:** United States

**ORCID number:** Megan Lynn Otte 0000-0003-2984-8198; Rizwan Ahmad 0000-0003-0331-6687; Punita Dhawan 0000-0003-3434-8155; Amar B Singh 0000-0003-1908-8594.

**Corresponding Author's Membership in Professional Societies:** American Gastrointestinal Association (AGA), 992160.

**S-Editor:** Liu GL

**L-Editor:** A

**P-Editor:** Liu GL

## REFERENCES

- 1 **Pariente B**, Hu S, Bettenworth D, Specia S, Desreumaux P, Meuwis MA, Danese S, Rieder F, Louis E. Treatments for Crohn's Disease-Associated Bowel Damage: A Systematic Review. *Clin Gastroenterol Hepatol* 2019; **17**: 847-856 [PMID: 30012430 DOI: 10.1016/j.cgh.2018.06.043]
- 2 **de Sablet T**, Potiron L, Marquis M, Bussi re FI, Lacroix-Lamand  S, Laurent F. Cryptosporidium parvum increases intestinal permeability through interaction with epithelial cells and IL-1  and TNF  released by inflammatory monocytes. *Cell Microbiol* 2016; **18**: 1871-1880 [PMID: 27324279 DOI: 10.1111/cmi.12632]
- 3 **Bernink JH**, Peters CP, Munneke M, te Velde AA, Meijer SL, Weijer K, Hreggvidsdottir HS, Heinsbroek SE, Legrand N, Buskens CJ, Bemelman WA, Mj sberg JM, Spits H. Human type 1 innate lymphoid cells accumulate in inflamed mucosal tissues. *Nat Immunol* 2013; **14**: 221-229 [PMID: 23334791 DOI: 10.1038/ni.2534]
- 4 **McDowell C**, Farooq U, Haseeb M. Inflammatory Bowel Disease. 2022 Jun 27. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2022 Jan- [PMID: 29262182]
- 5 **Fakhoury M**, Negrulj R, Mooranian A, Al-Salami H. Inflammatory bowel disease: clinical aspects and treatments. *J Inflamm Res* 2014; **7**: 113-120 [PMID: 25075198 DOI: 10.2147/JIR.S65979]
- 6 **Sadaghian Sadabad M**, Regeling A, de Goffau MC, Blokzijl T, Weersma RK, Penders J, Faber KN, Harmsen HJ, Dijkstra G. The ATG16L1-T300A allele impairs clearance of pathosymbionts in the inflamed ileal mucosa of Crohn's disease patients. *Gut* 2015; **64**: 1546-1552 [PMID: 25253126 DOI: 10.1136/gutjnl-2014-307289]
- 7 **Lavoie S**, Conway KL, Lassen KG, Jijon HB, Pan H, Chun E, Michaud M, Lang JK, Gallini Comeau CA, Dreyfuss JM, Glickman JN, Vlamakis H, Ananthakrishnan A, Kostic A, Garrett WS, Xavier RJ. The Crohn's disease polymorphism, ATG16L1 T300A, alters the gut microbiota and enhances the local Th1/Th17 response. *Elife* 2019; **8** [PMID: 30666959 DOI: 10.7554/eLife.39982]
- 8 **Sommer K**, Wendl M, M ller TM, Heidbreder K, Voskens C, Neurath MF, Zundler S. Intestinal Mucosal Wound Healing and Barrier Integrity in IBD-Crosstalk and Trafficking of Cellular Players. *Front Med (Lausanne)* 2021; **8**: 643973 [PMID: 33834033 DOI: 10.3389/fmed.2021.643973]
- 9 **Neurath MF**. Cytokines in inflammatory bowel disease. *Nat Rev Immunol* 2014; **14**: 329-342 [PMID: 24751956 DOI: 10.1038/nri3661]
- 10 **Fine S**, Papamichael K, Cheifetz AS. Etiology and Management of Lack or Loss of Response to Anti-Tumor Necrosis Factor Therapy in Patients With Inflammatory Bowel Disease. *Gastroenterol Hepatol (N Y)* 2019; **15**: 656-665 [PMID: 31892912]
- 11 **Neurath MF**. New targets for mucosal healing and therapy in inflammatory bowel diseases. *Mucosal Immunol* 2014; **7**: 6-19 [PMID: 24084775 DOI: 10.1038/mi.2013.73]

- 12 **Klenske E**, Bojarski C, Waldner M, Rath T, Neurath MF, Atreya R. Targeting mucosal healing in Crohn's disease: what the clinician needs to know. *Therap Adv Gastroenterol* 2019; **12**: 1756284819856865 [PMID: [31236140](#) DOI: [10.1177/1756284819856865](#)]
- 13 **Baars JE**, Vogelaar L, Wolfhagen FH, Biermann K, Kuipers EJ, van der Woude CJ. A short course of corticosteroids prior to surveillance colonoscopy to decrease mucosal inflammation in inflammatory bowel disease patients: results from a randomized controlled trial. *J Crohns Colitis* 2010; **4**: 661-668 [PMID: [21122577](#) DOI: [10.1016/j.crohns.2010.07.011](#)]
- 14 **Modigliani R**, Mary JY, Simon JF, Cortot A, Soule JC, Gendre JP, Rene E. Clinical, biological, and endoscopic picture of attacks of Crohn's disease. Evolution on prednisolone. Groupe d'Etude Thérapeutique des Affections Inflammatoires Digestives. *Gastroenterology* 1990; **98**: 811-818 [PMID: [2179031](#) DOI: [10.1016/0016-5085\(90\)90002-i](#)]
- 15 **Peyrin-Biroulet L**, Sandborn W, Sands BE, Reinisch W, Bemelman W, Bryant RV, D'Haens G, Dotan I, Dubinsky M, Feagan B, Fiorino G, Geary R, Krishnareddy S, Lakatos PL, Loftus EV Jr, Marteau P, Munkholm P, Murdosh TB, Ordás I, Panaccione R, Riddell RH, Ruel J, Rubin DT, Samaan M, Siegel CA, Silverberg MS, Stoker J, Schreiber S, Travis S, Van Assche G, Danese S, Panes J, Bouguen G, O'Donnell S, Pariente B, Winer S, Hanauer S, Colombel JF. Selecting Therapeutic Targets in Inflammatory Bowel Disease (STRIDE): Determining Therapeutic Goals for Treat-to-Target. *Am J Gastroenterol* 2015; **110**: 1324-1338 [PMID: [26303131](#) DOI: [10.1038/ajg.2015.233](#)]
- 16 **Wall CL**, Geary RB, Day AS. Treatment of Active Crohn's Disease with Exclusive and Partial Enteral Nutrition: A Pilot Study in Adults. *Inflamm Intest Dis* 2018; **2**: 219-227 [PMID: [30221149](#) DOI: [10.1159/000489630](#)]
- 17 **Horwat P**, Kopeć S, Garczyk A, Kaliciak I, Starega Z, Drogowski K, Mardas M, Stelmach-Mardas M. Influence of Enteral Nutrition on Gut Microbiota Composition in Patients with Crohn's Disease: A Systematic Review. *Nutrients* 2020; **12** [PMID: [32842543](#) DOI: [10.3390/nu12092551](#)]
- 18 **Römkens TE**, Kampschreur MT, Drenth JP, van Oijen MG, de Jong DJ. High mucosal healing rates in 5-ASA-treated ulcerative colitis patients: results of a meta-analysis of clinical trials. *Inflamm Bowel Dis* 2012; **18**: 2190-2198 [PMID: [22419617](#) DOI: [10.1002/ibd.22939](#)]
- 19 **Pearson DC**, May GR, Fick GH, Sutherland LR. Azathioprine and 6-mercaptopurine in Crohn disease. A meta-analysis. *Ann Intern Med* 1995; **123**: 132-142 [PMID: [7778826](#) DOI: [10.7326/0003-4819-123-2-199507150-00009](#)]
- 20 **Bunea MC**, Diculescu VC, Enculescu M, Iovu H, Enache TA. Redox Mechanism of Azathioprine and Its Interaction with DNA. *Int J Mol Sci* 2021; **22** [PMID: [34202734](#) DOI: [10.3390/ijms22136805](#)]
- 21 **Huang Z**, Chao K, Li M, Zhi M, Tang J, Hu P, Gao X. Methotrexate for Refractory Crohn's Disease Compared with Thiopurines: A Retrospective Non-head-to-head Controlled Study. *Inflamm Bowel Dis* 2017; **23**: 440-447 [PMID: [28129286](#) DOI: [10.1097/MIB.0000000000001022](#)]
- 22 **Colombel JF**, Sandborn WJ, Reinisch W, Mantzaris GJ, Kornbluth A, Rachmilewitz D, Lichtiger S, D'Haens G, Diamond RH, Broussard DL, Tang KL, van der Woude CJ, Rutgeerts P; SONIC Study Group. Infliximab, azathioprine, or combination therapy for Crohn's disease. *N Engl J Med* 2010; **362**: 1383-1395 [PMID: [20393175](#) DOI: [10.1056/NEJMoa0904492](#)]
- 23 **Christensen B**, Gibson PR, Micic D, Colman RJ, Goepfing SR, Kassim O, Yarur A, Weber CR, Cohen RD, Rubin DT. Safety and Efficacy of Combination Treatment With Calcineurin Inhibitors and Vedolizumab in Patients With Refractory Inflammatory Bowel Disease. *Clin Gastroenterol Hepatol* 2019; **17**: 486-493 [PMID: [29751166](#) DOI: [10.1016/j.cgh.2018.04.060](#)]
- 24 **Hanauer SB**, Sandborn WJ, Rutgeerts P, Fedorak RN, Lukas M, MacIntosh D, Panaccione R, Wolf D, Pollack P. Human anti-tumor necrosis factor monoclonal antibody (adalimumab) in Crohn's disease: the CLASSIC-I trial. *Gastroenterology* 2006; **130**: 323-33; quiz 591 [PMID: [16472588](#) DOI: [10.1053/j.gastro.2005.11.030](#)]
- 25 **Suenaert P**, Bulteel V, Lemmens L, Noman M, Geypens B, Van Assche G, Geboes K, Ceuppens JL, Rutgeerts P. Anti-tumor necrosis factor treatment restores the gut barrier in Crohn's disease. *Am J Gastroenterol* 2002; **97**: 2000-2004 [PMID: [12190167](#) DOI: [10.1111/j.1572-0241.2002.05914.x](#)]
- 26 **Baert FJ**, D'Haens GR, Peeters M, Hiele MI, Schaible TF, Shealy D, Geboes K, Rutgeerts PJ. Tumor necrosis factor alpha antibody (infliximab) therapy profoundly down-regulates the inflammation in Crohn's ileocolitis. *Gastroenterology* 1999; **116**: 22-28 [PMID: [9869598](#) DOI: [10.1016/s0016-5085\(99\)70224-6](#)]
- 27 **Rutgeerts P**, Vermeire S, Van Assche G. Mucosal healing in inflammatory bowel disease: impossible ideal or therapeutic target? *Gut* 2007; **56**: 453-455 [PMID: [17369375](#) DOI: [10.1136/gut.2005.088732](#)]
- 28 **Rutgeerts P**, Van Assche G, Sandborn WJ, Wolf DC, Geboes K, Colombel JF, Reinisch W; EXTEND Investigators, Kumar A, Lazar A, Camez A, Lomax KG, Pollack PF, D'Haens G. Adalimumab induces and maintains mucosal healing in patients with Crohn's disease: data from the EXTEND trial. *Gastroenterology* 2012; **142**: 1102-1111.e2 [PMID: [22326435](#) DOI: [10.1053/j.gastro.2012.01.035](#)]
- 29 **Schreiber S**, Rutgeerts P, Fedorak RN, Khaliq-Kareemi M, Kamm MA, Boivin M, Bernstein CN, Staun M, Thomsen OØ, Innes A; CDP870 Crohn's Disease Study Group. A randomized, placebo-controlled trial of certolizumab pegol (CDP870) for treatment of Crohn's disease. *Gastroenterology* 2005; **129**: 807-818 [PMID: [16143120](#) DOI: [10.1053/j.gastro.2005.06.064](#)]
- 30 **Rutgeerts P**, Gasink C, Chan D, Lang Y, Pollack P, Colombel JF, Wolf DC, Jacobstein D, Johans J, Szapary P, Adedokun OJ, Feagan BG, Sandborn WJ. Efficacy of Ustekinumab for Inducing Endoscopic Healing in Patients With Crohn's Disease. *Gastroenterology* 2018; **155**: 1045-1058 [PMID: [29909019](#) DOI: [10.1053/j.gastro.2018.06.035](#)]
- 31 **D'Haens G**, Panaccione R, Baert F, Bossuyt P, Colombel JF, Danese S, Dubinsky M, Feagan BG, Hisamatsu T, Lim A, Lindsay JO, Loftus EV Jr, Panés J, Peyrin-Biroulet L, Ran Z, Rubin DT, Sandborn WJ, Schreiber S, Neimark E, Song A, Kligys K, Pang Y, Pivorunas V, Berg S, Duan WR, Huang B, Kalabic J, Liao X, Robinson A, Wallace K, Ferrante M. Risankizumab as induction therapy for Crohn's disease: results from the phase 3 ADVANCE and MOTIVATE induction trials. *Lancet* 2022; **399**: 2015-2030 [PMID: [35644154](#) DOI: [10.1016/S0140-6736\(22\)00467-6](#)]
- 32 **Sakuraba A**, Annunziata ML, Cohen RD, Hanauer SB, Rubin DT. Mucosal healing is associated with improved long-term outcome of maintenance therapy with natalizumab in Crohn's disease. *Inflamm Bowel Dis* 2013; **19**: 2577-2583 [PMID: [24132161](#) DOI: [10.1097/MIB.0b013e3182a8df32](#)]
- 33 **Noman M**, Ferrante M, Bisschops R, De Hertogh G, Van den Broeck K, Rans K, Rutgeerts P, Vermeire S, Van Assche G.

- Vedolizumab Induces Long-term Mucosal Healing in Patients With Crohn's Disease and Ulcerative Colitis. *J Crohns Colitis* 2017; **11**: 1085-1089 [PMID: 28369329 DOI: 10.1093/ecco-jcc/jjx048]
- 34 **Feagan BG**, Rutgeerts P, Sands BE, Hanauer S, Colombel JF, Sandborn WJ, Van Assche G, Axler J, Kim HJ, Danese S, Fox I, Milch C, Sankoh S, Wyant T, Xu J, Parikh A; GEMINI 1 Study Group. Vedolizumab as induction and maintenance therapy for ulcerative colitis. *N Engl J Med* 2013; **369**: 699-710 [PMID: 23964932 DOI: 10.1056/NEJMoa1215734]
- 35 **Groschwitz KR**, Hogan SP. Intestinal barrier function: molecular regulation and disease pathogenesis. *J Allergy Clin Immunol* 2009; **124**: 3-20; quiz 21 [PMID: 19560575 DOI: 10.1016/j.jaci.2009.05.038]
- 36 **Blikslager AT**, Moeser AJ, Gookin JL, Jones SL, Odle J. Restoration of barrier function in injured intestinal mucosa. *Physiol Rev* 2007; **87**: 545-564 [PMID: 17429041 DOI: 10.1152/physrev.00012.2006]
- 37 **Rusu AD**, Georgiou M. The multifarious regulation of the apical junctional complex. *Open Biol* 2020; **10**: 190278 [PMID: 32070233 DOI: 10.1098/rsob.190278]
- 38 **Chelakkot C**, Ghim J, Ryu SH. Mechanisms regulating intestinal barrier integrity and its pathological implications. *Exp Mol Med* 2018; **50**: 1-9 [PMID: 30115904 DOI: 10.1038/s12276-018-0126-x]
- 39 **Buckley A**, Turner JR. Cell Biology of Tight Junction Barrier Regulation and Mucosal Disease. *Cold Spring Harb Perspect Biol* 2018; **10** [PMID: 28507021 DOI: 10.1101/cshperspect.a029314]
- 40 **Anderson JM**, Van Itallie CM. Physiology and function of the tight junction. *Cold Spring Harb Perspect Biol* 2009; **1**: a002584 [PMID: 20066090 DOI: 10.1101/cshperspect.a002584]
- 41 **Spadaro D**, Tapia R, Jond L, Sudol M, Fanning AS, Citi S. ZO proteins redundantly regulate the transcription factor DbpA/ZONAB. *J Biol Chem* 2014; **289**: 22500-22511 [PMID: 24986862 DOI: 10.1074/jbc.M114.556449]
- 42 **Balda MS**, Garrett MD, Matter K. The ZO-1-associated Y-box factor ZONAB regulates epithelial cell proliferation and cell density. *J Cell Biol* 2003; **160**: 423-432 [PMID: 12566432 DOI: 10.1083/jcb.200210020]
- 43 **Oka T**, Remue E, Meerschaert K, Vanloo B, Boucherie C, Gfeller D, Bader GD, Sidhu SS, Vandekerckhove J, Gettemans J, Sudol M. Functional complexes between YAP2 and ZO-2 are PDZ domain-dependent, and regulate YAP2 nuclear localization and signalling. *Biochem J* 2010; **432**: 461-472 [PMID: 20868367 DOI: 10.1042/BJ20100870]
- 44 **Fan S**, Smith MS, Keeney J, O'Leary MN, Nusrat A, Parkos CA. JAM-A signals through the Hippo pathway to regulate intestinal epithelial proliferation. *iScience* 2022; **25**: 104316 [PMID: 35602956 DOI: 10.1016/j.isci.2022.104316]
- 45 **Nava P**, Capaldo CT, Koch S, Kolegraff K, Rankin CR, Farkas AE, Feasel ME, Li L, Addis C, Parkos CA, Nusrat A. JAM-A regulates epithelial proliferation through Akt/ $\beta$ -catenin signalling. *EMBO Rep* 2011; **12**: 314-320 [PMID: 21372850 DOI: 10.1038/embor.2011.16]
- 46 **Kuo WT**, Shen L, Zuo L, Shashikanth N, Ong MLDM, Wu L, Zha J, Edelblum KL, Wang Y, Nilsen SP, Turner JR. Inflammation-induced Occludin Downregulation Limits Epithelial Apoptosis by Suppressing Caspase-3 Expression. *Gastroenterology* 2019; **157**: 1323-1337 [PMID: 31401143 DOI: 10.1053/j.gastro.2019.07.058]
- 47 **Chelakkot C**, Ghim J, Rajasekaran N, Choi JS, Kim JH, Jang MH, Shin YK, Suh PG, Ryu SH. Intestinal Epithelial Cell-Specific Deletion of PLD2 Alleviates DSS-Induced Colitis by Regulating Occludin. *Sci Rep* 2017; **7**: 1573 [PMID: 28484281 DOI: 10.1038/s41598-017-01797-y]
- 48 **Pope JL**, Ahmad R, Bhat AA, Washington MK, Singh AB, Dhawan P. Claudin-1 overexpression in intestinal epithelial cells enhances susceptibility to adenomatous polyposis coli-mediated colon tumorigenesis. *Mol Cancer* 2014; **13**: 167 [PMID: 24997475 DOI: 10.1186/1476-4598-13-167]
- 49 **Gowrikumar S**, Ahmad R, Uppada SB, Washington MK, Shi C, Singh AB, Dhawan P. Upregulated claudin-1 expression promotes colitis-associated cancer by promoting  $\beta$ -catenin phosphorylation and activation in Notch/p-AKT-dependent manner. *Oncogene* 2019; **38**: 5321-5337 [PMID: 30971761 DOI: 10.1038/s41388-019-0795-5]
- 50 **Ahmad R**, Kumar B, Pan K, Dhawan P, Singh AB. HDAC-4 regulates claudin-2 expression in EGFR-ERK1/2 dependent manner to regulate colonic epithelial cell differentiation. *Oncotarget* 2017; **8**: 87718-87736 [PMID: 29152115 DOI: 10.18632/oncotarget.21190]
- 51 **Mankertz J**, Hillenbrand B, Tavalali S, Huber O, Fromm M, Schulzke JD. Functional crosstalk between Wnt signaling and Cdx-related transcriptional activation in the regulation of the claudin-2 promoter activity. *Biochem Biophys Res Commun* 2004; **314**: 1001-1007 [PMID: 14751232 DOI: 10.1016/j.bbrc.2003.12.185]
- 52 **Zhang YG**, Wu S, Lu R, Zhou D, Zhou J, Carmeliet G, Petrof E, Claud EC, Sun J. Tight junction CLDN2 gene is a direct target of the vitamin D receptor. *Sci Rep* 2015; **5**: 10642 [PMID: 26212084 DOI: 10.1038/srep10642]
- 53 **Sonoda N**, Furuse M, Sasaki H, Yonemura S, Katahira J, Horiguchi Y, Tsukita S. Clostridium perfringens enterotoxin fragment removes specific claudins from tight junction strands: Evidence for direct involvement of claudins in tight junction barrier. *J Cell Biol* 1999; **147**: 195-204 [PMID: 10508866 DOI: 10.1083/jcb.147.1.195]
- 54 **Prasad S**, Mingrino R, Kaukinen K, Hayes KL, Powell RM, MacDonald TT, Collins JE. Inflammatory processes have differential effects on claudins 2, 3 and 4 in colonic epithelial cells. *Lab Invest* 2005; **85**: 1139-1162 [PMID: 16007110 DOI: 10.1038/Labinvest.3700316]
- 55 **Yuan B**, Zhou S, Lu Y, Liu J, Jin X, Wan H, Wang F. Changes in the Expression and Distribution of Claudins, Increased Epithelial Apoptosis, and a Mannan-Binding Lectin-Associated Immune Response Lead to Barrier Dysfunction in Dextran Sodium Sulfate-Induced Rat Colitis. *Gut Liver* 2015; **9**: 734-740 [PMID: 25717051 DOI: 10.5009/gnl14155]
- 56 **Wang K**, Ding Y, Xu C, Hao M, Li H, Ding L. Cldn-7 deficiency promotes experimental colitis and associated carcinogenesis by regulating intestinal epithelial integrity. *Oncoimmunology* 2021; **10**: 1923910 [PMID: 34026335 DOI: 10.1080/2162402X.2021.1923910]
- 57 **Martin-Rodriguez O**, Gauthier T, Bonnefoy F, Couturier M, Daoui A, Chagué C, Valmary-Degano S, Gay C, Saas P, Perruche S. Pro-Resolving Factors Released by Macrophages After Efferocytosis Promote Mucosal Wound Healing in Inflammatory Bowel Disease. *Front Immunol* 2021; **12**: 754475 [PMID: 35003066 DOI: 10.3389/fimmu.2021.754475]
- 58 **Xing T**, Benderman LJ, Sabu S, Parker J, Yang J, Lu Q, Ding L, Chen YH. Tight Junction Protein Claudin-7 Is Essential for Intestinal Epithelial Stem Cell Self-Renewal and Differentiation. *Cell Mol Gastroenterol Hepatol* 2020; **9**: 641-659 [PMID: 31874254 DOI: 10.1016/j.jcmgh.2019.12.005]
- 59 **Nübel T**, Preobraschenski J, Tuncay H, Weiss T, Kuhn S, Ladwein M, Langbein L, Zöller M. Claudin-7 regulates

- EpCAM-mediated functions in tumor progression. *Mol Cancer Res* 2009; **7**: 285-299 [PMID: 19276185 DOI: 10.1158/1541-7786.MCR-08-0200]
- 60 **Tamura A**, Kitano Y, Hata M, Katsuno T, Moriwaki K, Sasaki H, Hayashi H, Suzuki Y, Noda T, Furuse M, Tsukita S. Megaintestine in claudin-15-deficient mice. *Gastroenterology* 2008; **134**: 523-534 [PMID: 18242218 DOI: 10.1053/j.gastro.2007.11.040]
- 61 **Wei M**, Ma Y, Shen L, Xu Y, Liu L, Bu X, Guo Z, Qin H, Li Z, Wang Z, Wu K, Yao L, Li J, Zhang J. NDRG2 regulates adherens junction integrity to restrict colitis and tumorigenesis. *EBioMedicine* 2020; **61**: 103068 [PMID: 33099085 DOI: 10.1016/j.ebiom.2020.103068]
- 62 **Aban CE**, Lombardi A, Neiman G, Biani MC, La Greca A, Waisman A, Moro LN, Sevlever G, Miriuka S, Luzzani C. Downregulation of E-cadherin in pluripotent stem cells triggers partial EMT. *Sci Rep* 2021; **11**: 2048 [PMID: 33479502 DOI: 10.1038/s41598-021-81735-1]
- 63 **Silvestre J**, Kenis PJ, Leckband DE. Cadherin and integrin regulation of epithelial cell migration. *Langmuir* 2009; **25**: 10092-10099 [PMID: 19583181 DOI: 10.1021/la901109e]
- 64 **Bondow BJ**, Faber ML, Wojta KJ, Walker EM, Battle MA. E-cadherin is required for intestinal morphogenesis in the mouse. *Dev Biol* 2012; **371**: 1-12 [PMID: 22766025 DOI: 10.1016/j.ydbio.2012.06.005]
- 65 **Smalley-Freed WG**, Efimov A, Burnett PE, Short SP, Davis MA, Gumucio DL, Washington MK, Coffey RJ, Reynolds AB. p120-catenin is essential for maintenance of barrier function and intestinal homeostasis in mice. *J Clin Invest* 2010; **120**: 1824-1835 [PMID: 20484816 DOI: 10.1172/JCI41414]
- 66 **Schlegelmilch K**, Mohseni M, Kirak O, Pruszk J, Rodriguez JR, Zhou D, Kreger BT, Vasioukhin V, Avruch J, Brummelkamp TR, Camargo FD. Yap1 acts downstream of  $\alpha$ -catenin to control epidermal proliferation. *Cell* 2011; **144**: 782-795 [PMID: 21376238 DOI: 10.1016/j.cell.2011.02.031]
- 67 **Rima M**, Daghni M, Lopez A, Fajloun Z, Lefrancois L, Dunach M, Mori Y, Merle P, Brusés JL, De Waard M, Ronjat M. Down-regulation of the Wnt/ $\beta$ -catenin signaling pathway by Cacb4. *Mol Biol Cell* 2017; **28**: 3699-3708 [PMID: 29021340 DOI: 10.1091/mbc.E17-01-0076]
- 68 **Hwang SG**, Yu SS, Ryu JH, Jeon HB, Yoo YJ, Eom SH, Chun JS. Regulation of beta-catenin signaling and maintenance of chondrocyte differentiation by ubiquitin-independent proteasomal degradation of alpha-catenin. *J Biol Chem* 2005; **280**: 12758-12765 [PMID: 15695815 DOI: 10.1074/jbc.M413367200]
- 69 **Nishida A**, Inoue R, Inatomi O, Bamba S, Naito Y, Andoh A. Gut microbiota in the pathogenesis of inflammatory bowel disease. *Clin J Gastroenterol* 2018; **11**: 1-10 [PMID: 29285689 DOI: 10.1007/s12328-017-0813-5]
- 70 **Lavelle A**, Sokol H. Gut microbiota-derived metabolites as key actors in inflammatory bowel disease. *Nat Rev Gastroenterol Hepatol* 2020; **17**: 223-237 [PMID: 32076145 DOI: 10.1038/s41575-019-0258-z]
- 71 **Pickard JM**, Zeng MY, Caruso R, Núñez G. Gut microbiota: Role in pathogen colonization, immune responses, and inflammatory disease. *Immunol Rev* 2017; **279**: 70-89 [PMID: 28856738 DOI: 10.1111/immr.12567]
- 72 **Ni J**, Wu GD, Albenberg L, Tomov VT. Gut microbiota and IBD: causation or correlation? *Nat Rev Gastroenterol Hepatol* 2017; **14**: 573-584 [PMID: 28743984 DOI: 10.1038/nrgastro.2017.88]
- 73 **Al Bander Z**, Nitert MD, Mousa A, Naderpoor N. The Gut Microbiota and Inflammation: An Overview. *Int J Environ Res Public Health* 2020; **17** [PMID: 33086688 DOI: 10.3390/ijerph17207618]
- 74 **Sankarasubramanian J**, Ahmad R, Avuthu N, Singh AB, Guda C. Gut Microbiota and Metabolic Specificity in Ulcerative Colitis and Crohn's Disease. *Front Med (Lausanne)* 2020; **7**: 606298 [PMID: 33330572 DOI: 10.3389/fmed.2020.606298]
- 75 **Lama Tamang R**, Juritsch AF, Ahmad R, Salomon JD, Dhawan P, Ramer-Tait AE, Singh AB. The diet-microbiota axis: a key regulator of intestinal permeability in human health and disease. *Tissue Barriers* 2022; 2077069 [PMID: 35603609 DOI: 10.1080/21688370.2022.2077069]
- 76 **Babbini BA**, Sasaki M, Gerner-Schmidt KW, Nusrat A, Klapproth JM. The bacterial virulence factor lymphostatin compromises intestinal epithelial barrier function by modulating rho GTPases. *Am J Pathol* 2009; **174**: 1347-1357 [PMID: 19286565 DOI: 10.2353/ajpath.2009.080640]
- 77 **Li Q**, Zhang Q, Wang C, Li N, Li J. Invasion of enteropathogenic Escherichia coli into host cells through epithelial tight junctions. *FEBS J* 2008; **275**: 6022-6032 [PMID: 19016848 DOI: 10.1111/j.1742-4658.2008.06731.x]
- 78 **Jepson MA**, Schlecht HB, Collares-Buzato CB. Localization of dysfunctional tight junctions in Salmonella enterica serovar typhimurium-infected epithelial layers. *Infect Immun* 2000; **68**: 7202-7208 [PMID: 11083857 DOI: 10.1128/IAI.68.12.7202-7208.2000]
- 79 **Chen ML**, Ge Z, Fox JG, Schauer DB. Disruption of tight junctions and induction of proinflammatory cytokine responses in colonic epithelial cells by Campylobacter jejuni. *Infect Immun* 2006; **74**: 6581-6589 [PMID: 17015453 DOI: 10.1128/IAI.00958-06]
- 80 **Wu S**, Rhee KJ, Zhang M, Franco A, Sears CL. Bacteroides fragilis toxin stimulates intestinal epithelial cell shedding and gamma-secretase-dependent E-cadherin cleavage. *J Cell Sci* 2007; **120**: 1944-1952 [PMID: 17504810 DOI: 10.1242/jcs.03455]
- 81 **Abrams GD**, Bauer H, Sprinz H. Influence of the normal flora on mucosal morphology and cellular renewal in the ileum. A comparison of germ-free and conventional mice. *Lab Invest* 1963; **12**: 355-364 [PMID: 14010768]
- 82 **Pull SL**, Doherty JM, Mills JC, Gordon JI, Stappenbeck TS. Activated macrophages are an adaptive element of the colonic epithelial progenitor niche necessary for regenerative responses to injury. *Proc Natl Acad Sci U S A* 2005; **102**: 99-104 [PMID: 15615857 DOI: 10.1073/pnas.0405979102]
- 83 **Alam A**, Leoni G, Wentworth CC, Kwal JM, Wu H, Ardita CS, Swanson PA, Lambeth JD, Jones RM, Nusrat A, Neish AS. Redox signaling regulates commensal-mediated mucosal homeostasis and restitution and requires formyl peptide receptor 1. *Mucosal Immunol* 2014; **7**: 645-655 [PMID: 24192910 DOI: 10.1038/mi.2013.84]
- 84 **Leoni G**, Alam A, Neumann PA, Lambeth JD, Cheng G, McCoy J, Hilgarth RS, Kundu K, Murthy N, Kusters D, Reutelingperger C, Perretti M, Parkos CA, Neish AS, Nusrat A. Annexin A1, formyl peptide receptor, and NOX1 orchestrate epithelial repair. *J Clin Invest* 2013; **123**: 443-454 [PMID: 23241962 DOI: 10.1172/JCI65831]
- 85 **Swanson PA 2nd**, Kumar A, Samarín S, Vijay-Kumar M, Kundu K, Murthy N, Hansen J, Nusrat A, Neish AS. Enteric

- commensal bacteria potentiate epithelial restitution *via* reactive oxygen species-mediated inactivation of focal adhesion kinase phosphatases. *Proc Natl Acad Sci U S A* 2011; **108**: 8803-8808 [PMID: 21555563 DOI: 10.1073/pnas.1010042108]
- 86 **Alam A**, Leoni G, Quiros M, Wu H, Desai C, Nishio H, Jones RM, Nusrat A, Neish AS. The microenvironment of injured murine gut elicits a local pro-restitutive microbiota. *Nat Microbiol* 2016; **1**: 15021 [PMID: 27571978 DOI: 10.1038/nmicrobiol.2015.21]
- 87 **Liang L**, Liu L, Zhou W, Yang C, Mai G, Li H, Chen Y. Gut microbiota-derived butyrate regulates gut mucus barrier repair by activating the macrophage/WNT/ERK signaling pathway. *Clin Sci (Lond)* 2022; **136**: 291-307 [PMID: 35194640 DOI: 10.1042/CS20210778]
- 88 **Lee JS**, Wang RX, Goldberg MS, Clifford GP, Kao DJ, Colgan SP. Microbiota-Sourced Purines Support Wound Healing and Mucous Barrier Function. *iScience* 2020; **23**: 101226 [PMID: 32554188 DOI: 10.1016/j.isci.2020.101226]
- 89 **Cummings JH**, Macfarlane GT. The control and consequences of bacterial fermentation in the human colon. *J Appl Bacteriol* 1991; **70**: 443-459 [PMID: 1938669 DOI: 10.1111/j.1365-2672.1991.tb02739.x]
- 90 **Clausen MR**, Mortensen PB. Kinetic studies on colonocyte metabolism of short chain fatty acids and glucose in ulcerative colitis. *Gut* 1995; **37**: 684-689 [PMID: 8549946 DOI: 10.1136/gut.37.5.684]
- 91 **Jørgensen JR**, Clausen MR, Mortensen PB. Oxidation of short and medium chain C2-C8 fatty acids in Sprague-Dawley rat colonocytes. *Gut* 1997; **40**: 400-405 [PMID: 9135532 DOI: 10.1136/gut.40.3.400]
- 92 **Roediger WE**. Utilization of nutrients by isolated epithelial cells of the rat colon. *Gastroenterology* 1982; **83**: 424-429 [PMID: 7084619]
- 93 **Louis P**, Young P, Holtrop G, Flint HJ. Diversity of human colonic butyrate-producing bacteria revealed by analysis of the butyryl-CoA:acetate CoA-transferase gene. *Environ Microbiol* 2010; **12**: 304-314 [PMID: 19807780 DOI: 10.1111/j.1462-2920.2009.02066.x]
- 94 **Martens EC**, Lowe EC, Chiang H, Pudlo NA, Wu M, McNulty NP, Abbott DW, Henrissat B, Gilbert HJ, Bolam DN, Gordon JI. Recognition and degradation of plant cell wall polysaccharides by two human gut symbionts. *PLoS Biol* 2011; **9**: e1001221 [PMID: 22205877 DOI: 10.1371/journal.pbio.1001221]
- 95 **Tan H**, Zhao J, Zhang H, Zhai Q, Chen W. Novel strains of *Bacteroides fragilis* and *Bacteroides ovatus* alleviate the LPS-induced inflammation in mice. *Appl Microbiol Biotechnol* 2019; **103**: 2353-2365 [PMID: 30666361 DOI: 10.1007/s00253-019-09617-1]
- 96 **Ihekweazu FD**, Engevik MA, Ruan W, Shi Z, Fultz R, Engevik KA, Chang-Graham AL, Freeborn J, Park ES, Venable S, Horvath TD, Haidacher SJ, Haag AM, Goodwin A, Schady DA, Hyser JM, Spinler JK, Liu Y, Versalovic J. *Bacteroides ovatus* Promotes IL-22 Production and Reduces Trinitrobenzene Sulfonic Acid-Driven Colonic Inflammation. *Am J Pathol* 2021; **191**: 704-719 [PMID: 33516788 DOI: 10.1016/j.ajpath.2021.01.009]
- 97 **Negi S**, Saini S, Tandel N, Sahu K, Mishra RPN, Tyagi RK. Translating Treg Therapy for Inflammatory Bowel Disease in Humanized Mice. *Cells* 2021; **10** [PMID: 34440615 DOI: 10.3390/cells10081847]
- 98 **Park J**, Kim M, Kang SG, Jannasch AH, Cooper B, Patterson J, Kim CH. Short-chain fatty acids induce both effector and regulatory T cells by suppression of histone deacetylases and regulation of the mTOR-S6K pathway. *Mucosal Immunol* 2015; **8**: 80-93 [PMID: 24917457 DOI: 10.1038/mi.2014.44]
- 99 **Smith PM**, Howitt MR, Panikov N, Michaud M, Gallini CA, Bohlooly-Y M, Glickman JN, Garrett WS. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science* 2013; **341**: 569-573 [PMID: 23828891 DOI: 10.1126/science.1241165]
- 100 **Sun M**, Wu W, Chen L, Yang W, Huang X, Ma C, Chen F, Xiao Y, Zhao Y, Yao S, Carpio VH, Dann SM, Zhao Q, Liu Z, Cong Y. Microbiota-derived short-chain fatty acids promote Th1 cell IL-10 production to maintain intestinal homeostasis. *Nat Commun* 2018; **9**: 3555 [PMID: 30177845 DOI: 10.1038/s41467-018-05901-2]
- 101 **Pickert G**, Neufert C, Leppkes M, Zheng Y, Wittkopf N, Warntjen M, Lehr HA, Hirth S, Weigmann B, Wirtz S, Ouyang W, Neurath MF, Becker C. STAT3 Links IL-22 signaling in intestinal epithelial cells to mucosal wound healing. *J Exp Med* 2009; **206**: 1465-1472 [PMID: 19564350 DOI: 10.1084/jem.20082683]
- 102 **Ivanov II**, Atarashi K, Manel N, Brodie EL, Shima T, Karaoz U, Wei D, Goldfarb KC, Santee CA, Lynch SV, Tanoue T, Imaoka A, Itoh K, Takeda K, Umesaki Y, Honda K, Littman DR. Induction of intestinal Th17 cells by segmented filamentous bacteria. *Cell* 2009; **139**: 485-498 [PMID: 19836068 DOI: 10.1016/j.cell.2009.09.033]
- 103 **Flannigan KL**, Denning TL. Segmented filamentous bacteria-induced immune responses: a balancing act between host protection and autoimmunity. *Immunology* 2018; **154**: 537-546 [PMID: 29771448 DOI: 10.1111/imm.12950]
- 104 **Sugimoto K**, Ogawa A, Mizoguchi E, Shimomura Y, Andoh A, Bhan AK, Blumberg RS, Xavier RJ, Mizoguchi A. IL-22 ameliorates intestinal inflammation in a mouse model of ulcerative colitis. *J Clin Invest* 2008; **118**: 534-544 [PMID: 18172556 DOI: 10.1172/JCI33194]
- 105 **Shih VF**, Cox J, Kljavin NM, Dengler HS, Reichelt M, Kumar P, Rangell L, Kolls JK, Diehl L, Ouyang W, Ghilardi N. Homeostatic IL-23 receptor signaling limits Th17 response through IL-22-mediated containment of commensal microbiota. *Proc Natl Acad Sci U S A* 2014; **111**: 13942-13947 [PMID: 25201978 DOI: 10.1073/pnas.1323852111]
- 106 **Nastasi C**, Candela M, Bonefeld CM, Geisler C, Hansen M, Krejsgaard T, Biagi E, Andersen MH, Brigidi P, Ødum N, Litman T, Woetmann A. The effect of short-chain fatty acids on human monocyte-derived dendritic cells. *Sci Rep* 2015; **5**: 16148 [PMID: 26541096 DOI: 10.1038/srep16148]
- 107 **Millard AL**, Mertes PM, Ittelet D, Villard F, Jeannesson P, Bernard J. Butyrate affects differentiation, maturation and function of human monocyte-derived dendritic cells and macrophages. *Clin Exp Immunol* 2002; **130**: 245-255 [PMID: 12390312 DOI: 10.1046/j.0009-9104.2002.01977.x]
- 108 **Donohoe DR**, Garge N, Zhang X, Sun W, O'Connell TM, Bunger MK, Bultman SJ. The microbiome and butyrate regulate energy metabolism and autophagy in the mammalian colon. *Cell Metab* 2011; **13**: 517-526 [PMID: 21531334 DOI: 10.1016/j.cmet.2011.02.018]
- 109 **Tsuboi K**, Nishitani M, Takakura A, Imai Y, Komatsu M, Kawashima H. Autophagy Protects against Colitis by the Maintenance of Normal Gut Microflora and Secretion of Mucus. *J Biol Chem* 2015; **290**: 20511-20526 [PMID: 26149685 DOI: 10.1074/jbc.M114.632257]

- 110 **Yang L**, Liu C, Zhao W, He C, Ding J, Dai R, Xu K, Xiao L, Luo L, Liu S, Li W, Meng H. Impaired Autophagy in Intestinal Epithelial Cells Alters Gut Microbiota and Host Immune Responses. *Appl Environ Microbiol* 2018; **84** [PMID: 30006408 DOI: 10.1128/AEM.00880-18]
- 111 **Asano J**, Sato T, Ichinose S, Kajita M, Onai N, Shimizu S, Ohteki T. Intrinsic Autophagy Is Required for the Maintenance of Intestinal Stem Cells and for Irradiation-Induced Intestinal Regeneration. *Cell Rep* 2017; **20**: 1050-1060 [PMID: 28768191 DOI: 10.1016/j.celrep.2017.07.019]
- 112 **Trentesaux C**, Fraudeau M, Pitasi CL, Lemarchand J, Jacques S, Duche A, Letourneur F, Naser E, Bailly K, Schmitt A, Perret C, Romagnolo B. Essential role for autophagy protein ATG7 in the maintenance of intestinal stem cell integrity. *Proc Natl Acad Sci U S A* 2020; **117**: 11136-11146 [PMID: 32371487 DOI: 10.1073/pnas.1917174117]
- 113 **Amulic B**, Cazalet C, Hayes GL, Metzler KD, Zychlinsky A. Neutrophil function: from mechanisms to disease. *Annu Rev Immunol* 2012; **30**: 459-489 [PMID: 22224774 DOI: 10.1146/annurev-immunol-020711-074942]
- 114 **Zhang D**, Frenette PS. Cross talk between neutrophils and the microbiota. *Blood* 2019; **133**: 2168-2177 [PMID: 30898860 DOI: 10.1182/blood-2018-11-844555]
- 115 **Gorjifard S**, Goldszmid RS. Microbiota-myeloid cell crosstalk beyond the gut. *J Leukoc Biol* 2016; **100**: 865-879 [PMID: 27605211 DOI: 10.1189/jlb.3RI0516-222R]
- 116 **Rose WA 2nd**, Sakamoto K, Leifer CA. TLR9 is important for protection against intestinal damage and for intestinal repair. *Sci Rep* 2012; **2**: 574 [PMID: 22893852 DOI: 10.1038/srep00574]
- 117 **Rakoff-Nahoum S**, Paglino J, Eslami-Varzaneh F, Edberg S, Medzhitov R. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell* 2004; **118**: 229-241 [PMID: 15260992 DOI: 10.1016/j.cell.2004.07.002]
- 118 **Manieri NA**, Drylewicz MR, Miyoshi H, Stappenbeck TS. Igf2bp1 is required for full induction of Ptg2 mRNA in colonic mesenchymal stem cells in mice. *Gastroenterology* 2012; **143**: 110-21.e10 [PMID: 22465430 DOI: 10.1053/j.gastro.2012.03.037]
- 119 **Miyoshi H**, VanDussen KL, Malvin NP, Ryu SH, Wang Y, Sonnek NM, Lai CW, Stappenbeck TS. Prostaglandin E2 promotes intestinal repair through an adaptive cellular response of the epithelium. *EMBO J* 2017; **36**: 5-24 [PMID: 27797821 DOI: 10.15252/embj.201694660]
- 120 **Jain U**, Lai CW, Xiong S, Goodwin VM, Lu Q, Muegge BD, Christophi GP, VanDussen KL, Cummings BP, Young E, Hambor J, Stappenbeck TS. Temporal Regulation of the Bacterial Metabolite Deoxycholate during Colonic Repair Is Critical for Crypt Regeneration. *Cell Host Microbe* 2018; **24**: 353-363.e5 [PMID: 30122655 DOI: 10.1016/j.chom.2018.07.019]
- 121 **Butzner JD**, Parmar R, Bell CJ, Dalal V. Butyrate enema therapy stimulates mucosal repair in experimental colitis in the rat. *Gut* 1996; **38**: 568-573 [PMID: 8707089 DOI: 10.1136/gut.38.4.568]
- 122 **Oliva S**, Di Nardo G, Ferrari F, Mallardo S, Rossi P, Patrizi G, Cucchiara S, Stronati L. Randomised clinical trial: the effectiveness of *Lactobacillus reuteri* ATCC 55730 rectal enema in children with active distal ulcerative colitis. *Aliment Pharmacol Ther* 2012; **35**: 327-334 [PMID: 22150569 DOI: 10.1111/j.1365-2036.2011.04939.x]
- 123 **Wang L**, Cao H, Liu L, Wang B, Walker WA, Acra SA, Yan F. Activation of epidermal growth factor receptor mediates mucin production stimulated by p40, a *Lactobacillus rhamnosus* GG-derived protein. *J Biol Chem* 2014; **289**: 20234-20244 [PMID: 24895124 DOI: 10.1074/jbc.M114.553800]
- 124 **Patel RM**, Myers LS, Kurundkar AR, Maheshwari A, Nusrat A, Lin PW. Probiotic bacteria induce maturation of intestinal claudin 3 expression and barrier function. *Am J Pathol* 2012; **180**: 626-635 [PMID: 22155109 DOI: 10.1016/j.ajpath.2011.10.025]
- 125 **Carlsson AH**, Yakymenko O, Olivier I, Håkansson F, Postma E, Keita AV, Söderholm JD. Faecalibacterium prausnitzii supernatant improves intestinal barrier function in mice DSS colitis. *Scand J Gastroenterol* 2013; **48**: 1136-1144 [PMID: 23971882 DOI: 10.3109/00365521.2013.828773]
- 126 **Sokol H**, Pigneur B, Watterlot L, Lakhdari O, Bermúdez-Humarán LG, Gratadoux JJ, Blugeon S, Bridonneau C, Furet JP, Corthier G, Grangette C, Vasquez N, Pochart P, Trugnan G, Thomas G, Blottière HM, Doré J, Marteau P, Seksik P, Langella P. Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. *Proc Natl Acad Sci U S A* 2008; **105**: 16731-16736 [PMID: 18936492 DOI: 10.1073/pnas.0804812105]
- 127 **Dai C**, Zhao DH, Jiang M. VSL#3 probiotics regulate the intestinal epithelial barrier *in vivo* and *in vitro* via the p38 and ERK signaling pathways. *Int J Mol Med* 2012; **29**: 202-208 [PMID: 22089663 DOI: 10.3892/ijmm.2011.839]
- 128 **Grivennikov SI**, Tumanov AV, Liepinsh DJ, Kruglov AA, Marakusha BI, Shakhov AN, Murakami T, Drutskaya LN, Förster I, Clausen BE, Tessarollo L, Ryffel B, Kuprash DV, Nedospasov SA. Distinct and nonredundant *in vivo* functions of TNF produced by T cells and macrophages/neutrophils: protective and deleterious effects. *Immunity* 2005; **22**: 93-104 [PMID: 15664162 DOI: 10.1016/j.immuni.2004.11.016]
- 129 **Kojouharoff G**, Hans W, Obermeier F, Männel DN, Andus T, Schölmerich J, Gross V, Falk W. Neutralization of tumour necrosis factor (TNF) but not of IL-1 reduces inflammation in chronic dextran sulphate sodium-induced colitis in mice. *Clin Exp Immunol* 1997; **107**: 353-358 [PMID: 9030875 DOI: 10.1111/j.1365-2249.1997.291-ce1184.x]
- 130 **Semin I**, Ninnemann J, Bondareva M, Gimaev I, Kruglov AA. Interplay Between Microbiota, Toll-Like Receptors and Cytokines for the Maintenance of Epithelial Barrier Integrity. *Front Med (Lausanne)* 2021; **8**: 644333 [PMID: 34124086 DOI: 10.3389/fmed.2021.644333]
- 131 **Nosbaum A**, Prevel N, Truong HA, Mehta P, Ettinger M, Scharschmidt TC, Ali NH, Pauli ML, Abbas AK, Rosenblum MD. Cutting Edge: Regulatory T Cells Facilitate Cutaneous Wound Healing. *J Immunol* 2016; **196**: 2010-2014 [PMID: 26826250 DOI: 10.4049/jimmunol.1502139]
- 132 **Kuhn KA**, Schulz HM, Regner EH, Severs EL, Hendrickson JD, Mehta G, Whitney AK, Ir D, Ohri N, Robertson CE, Frank DN, Campbell EL, Colgan SP. Bacteroidales recruit IL-6-producing intraepithelial lymphocytes in the colon to promote barrier integrity. *Mucosal Immunol* 2018; **11**: 357-368 [PMID: 28812548 DOI: 10.1038/mi.2017.55]
- 133 **Madara JL**, Stafford J. Interferon-gamma directly affects barrier function of cultured intestinal epithelial monolayers. *J Clin Invest* 1989; **83**: 724-727 [PMID: 2492310 DOI: 10.1172/JCI113938]

- 134 **Suzuki T**, Yoshinaga N, Tanabe S. Interleukin-6 (IL-6) regulates claudin-2 expression and tight junction permeability in intestinal epithelium. *J Biol Chem* 2011; **286**: 31263-31271 [PMID: [21771795](#) DOI: [10.1074/jbc.M111.238147](#)]
- 135 **Kusugami K**, Fukatsu A, Tanimoto M, Shinoda M, Haruta J, Kuroiwa A, Ina K, Kanayama K, Ando T, Matsuura T. Elevation of interleukin-6 in inflammatory bowel disease is macrophage- and epithelial cell-dependent. *Dig Dis Sci* 1995; **40**: 949-959 [PMID: [7729284](#) DOI: [10.1007/BF02064182](#)]
- 136 **Danese S**, Vermeire S, Hellstern P, Panaccione R, Rogler G, Fraser G, Kohn A, Desreumaux P, Leong RW, Comer GM, Cataldi F, Banerjee A, Maguire MK, Li C, Rath N, Beebe J, Schreiber S. Randomised trial and open-label extension study of an anti-interleukin-6 antibody in Crohn's disease (ANDANTE I and II). *Gut* 2019; **68**: 40-48 [PMID: [29247068](#) DOI: [10.1136/gutjnl-2017-314562](#)]
- 137 **Beck PL**, Rosenberg IM, Xavier RJ, Koh T, Wong JF, Podolsky DK. Transforming growth factor-beta mediates intestinal healing and susceptibility to injury *in vitro* and *in vivo* through epithelial cells. *Am J Pathol* 2003; **162**: 597-608 [PMID: [12547717](#) DOI: [10.1016/s0002-9440\(10\)63853-9](#)]
- 138 **Bonnefoy F**, Gauthier T, Vallion R, Martin-Rodriguez O, Missey A, Daoui A, Valmary-Degano S, Saas P, Couturier M, Perruche S. Factors Produced by Macrophages Eliminating Apoptotic Cells Demonstrate Pro-Resolutive Properties and Terminate Ongoing Inflammation. *Front Immunol* 2018; **9**: 2586 [PMID: [30542342](#) DOI: [10.3389/fimmu.2018.02586](#)]
- 139 **Quiros M**, Nishio H, Neumann PA, Siuda D, Brazil JC, Azcutia V, Hilgarth R, O'Leary MN, Garcia-Hernandez V, Leoni G, Feng M, Bernal G, Williams H, Dedhia PH, Gerner-Smidt C, Spence J, Parkos CA, Denning TL, Nusrat A. Macrophage-derived IL-10 mediates mucosal repair by epithelial WISP-1 signaling. *J Clin Invest* 2017; **127**: 3510-3520 [PMID: [28783045](#) DOI: [10.1172/JCI90229](#)]
- 140 **Kühn R**, Löhler J, Rennick D, Rajewsky K, Müller W. Interleukin-10-deficient mice develop chronic enterocolitis. *Cell* 1993; **75**: 263-274 [PMID: [8402911](#) DOI: [10.1016/0092-8674\(93\)80068-p](#)]
- 141 **Song X**, Dai D, He X, Zhu S, Yao Y, Gao H, Wang J, Qu F, Qiu J, Wang H, Li X, Shen N, Qian Y. Growth Factor FGF2 Cooperates with Interleukin-17 to Repair Intestinal Epithelial Damage. *Immunity* 2015; **43**: 488-501 [PMID: [26320657](#) DOI: [10.1016/j.immuni.2015.06.024](#)]
- 142 **Rutz S**, Eidenschenk C, Ouyang W. IL-22, not simply a Th17 cytokine. *Immunol Rev* 2013; **252**: 116-132 [PMID: [23405899](#) DOI: [10.1111/imr.12027](#)]
- 143 **Hanash AM**, Dudakov JA, Hua G, O'Connor MH, Young LF, Singer NV, West ML, Jenq RR, Holland AM, Kappel LW, Ghosh A, Tsai JJ, Rao UK, Yim NL, Smith OM, Velardi E, Hawryluk EB, Murphy GF, Liu C, Fouser LA, Kolesnick R, Blazar BR, van den Brink MR. Interleukin-22 protects intestinal stem cells from immune-mediated tissue damage and regulates sensitivity to graft versus host disease. *Immunity* 2012; **37**: 339-350 [PMID: [22921121](#) DOI: [10.1016/j.immuni.2012.05.028](#)]
- 144 **Zenewicz LA**, Yancopoulos GD, Valenzuela DM, Murphy AJ, Stevens S, Flavell RA. Innate and adaptive interleukin-22 protects mice from inflammatory bowel disease. *Immunity* 2008; **29**: 947-957 [PMID: [19100701](#) DOI: [10.1016/j.immuni.2008.11.003](#)]
- 145 **Lindemans CA**, Calafiore M, Mertelsmann AM, O'Connor MH, Dudakov JA, Jenq RR, Velardi E, Young LF, Smith OM, Lawrence G, Ivanov JA, Fu YY, Takashima S, Hua G, Martin ML, O'Rourke KP, Lo YH, Mokry M, Romera-Hernandez M, Cupedo T, Dow L, Nieuwenhuis EE, Shroyer NF, Liu C, Kolesnick R, van den Brink MRM, Hanash AM. Interleukin-22 promotes intestinal-stem-cell-mediated epithelial regeneration. *Nature* 2015; **528**: 560-564 [PMID: [26649819](#) DOI: [10.1038/nature16460](#)]





Published by **Baishideng Publishing Group Inc**  
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

**Telephone:** +1-925-3991568

**E-mail:** [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

**Help Desk:** <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

